

Incidence and Plant stage relationship of rootrot incited by *Macrophomina phaseolina* on mothbean assessed using Novel Inoculation methods.

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Abstract

The ability of *Macrophomina phaseolina*, the casual agent of Moth bean to produce pycnidiospore in moth was studied in the field and under laboratory condition. The Root rot of moth bean (*Vigna aconitifolia* (Jacq.) Marechal, caused by *M. phaseolina*, is quite prevalent in the moth growing areas of Rajasthan and Uttar Pradesh state. The pathogen infects the moth plant at all ages and it results in a huge loss. The present study was undertaken to study the development of necrosis and occurrence of sclerotia and pycnidiospores in order to study the pathogenic variability of *M. phaseolina*. and to determine the morphological and pathogenic variability and their correlation with the age of vigna plant. The isolates showed variation in mycelia growth and sporulation. Variability is the very basis of survival of the pathogen and It was observed that the sclerotia were produced in collar regions of 15,30,45 and 60 day old plants but symptoms were found to be more prevalent at maturity stage as compared to initial, seedling and flowering stages. Data reported here indicates that the sclerotia contribute to death of infected plants.. It was observed that maximum disease incidence in plants occurs at maturity stage and susceptibility of plants to *Macrophomina* increased with age.

Key words: *Macrophomina phaseolina*, root rot, moth bean, sclerotia.

Introduction

Moth bean (*Vigna aconitifolia*(Jacq.) Marechal, is one of the important crops extensively grown in India mainly as a rain fed crop in kharif season primarily in the state of U.P. and Rajasthan besides other states on a limited scale. It belongs to the family 'Leguminosae'. (Jain and Mehra, 1980; Banerjee et al, 1983; Pant, 1990; Stephan, 1994; Chaturvedi. D., 2009). *Macrophomina phaseolina*(Tassi.) Goid. *Rhizoctonia bataticola*(Taub.) Butl.. causes number of diseases in legumes. *M. phaseolina* is universally distributed, fungi with high competitive ability, high lethal viability and a wide host range that makes the fungus economically important. Complex disease syndromes associated with this fungus are root rot, seed rot, seedling blight and charcoal rot. (Khare et al., 1973; Nene, 1977; Sharma and Gupta, 1981; Singh and Srivastava, 1989; Rout and Ingle, 1989; Pal, 1998; Lodha 1998; Rathore, 2000.) Number of fungal diseases of moth bean which

resulted in heavy losses of the crop was reported from various part of the country and the pathogen being both seed and soil borne may attack the plant at any growth stage, however maximum damage reported to be caused at seedling and seed maturing stage of the crop. (Aghnihotri et al., 1987; Byadgi and Hedge, 1988; Rathore et al, 1999, Lakhey and Shakya; 2007). The extent of yield loss depends on the stage at which the plants are affected. As in case of *M. phaseolina* it was observed plant was found to be Maximum diseased at Harvesting and Post Harvesting stages. Little work has been done on the pathogenic variability of *M. phaseolina*, and its correlation with plant age stages. Hence, these studies were undertaken.

Material and Methods

Pathogen Isolates:

Root rot affected plant samples were collected from moth growing fields. *M. phaseolina* (Tassi.) Goid. was isolated from these samples and single spores of each isolate were transferred to Sabarouds agar medium and incubated at $28 \pm 1^\circ\text{C}$.

Cultural and Morphological Variation:

Mycelial bits of 5 mm diameter were resumed from 7 day old culture of each isolate, inoculated on Sabarouds agar and incubated at $28 \pm 2^\circ\text{C}$ for 7 days. The morphological and cultural characteristics of colony were assessed for mycelia growth, sporulation, and sclerotia size.

Standardization of inoculation technique for pathogenic variation Studies :

Three methods of inoculation viz., Wound method, Soil drenching with inoculums and soil infestation were conducted.

In Wound Method, Moth bean (Jacq.) Marechal seedlings were planted in four wooden boxes (3 planted in each and was maintained on PDA (Potato Dextrose Agar). Plants were wounded above the soil line with a sterile scalpel. A small portion of mycelium from a 7 day old moth bean broth culture grown at $25 \pm 3^\circ\text{C}$ was placed into the wound cut of half of the plants. Then all wounds were covered with petroleum jelly (Vaseline) to prevent drying. After 14 days, lesions observations were made, then plants (according to their age) were cut off at the soil line, and observations on necrosis were made. (Upadhyay and Pandey, 2000). The collar regions and roots were examined under a dissecting microscope. Four wooden boxes with three plants in each were wound-inoculated with *Macrophomina phaseolina* as described above. Four boxes (three plants in each) wounded, but non-inoculated, plants served as controls.

In soil drenching method, seedlings of 8 days old were used. A spore suspension was poured around the root zone in the soil by disturbing the root zone.

In soil infestation method, the fungus was grown on autoclaved sorghum seeds for 8-10 days. The inoculums was mixed with soil and left for 7 days.

Then moth seeds were sown in captan solution (0.2%) for 2 hours and then sown.

When individual plants first expressed symptoms (7-14 days after inoculation). They were sectioned with a rotary microtome and stained with safranin. Sections were examined under a light microscope for the presence of *M. phaseolina* mycelium, Sclerotia, Pycnidiospores. (Dhingra and Sinclair, 1975.; Singh and Mehrotra, 1982, Mukherjee et al., 1983; Singh and Sandhu, 1994).

Results

The isolates showed great variability in colony characteristics such as mycelia growth, Sporulation.

Standardization of inoculation techniques for pathogenic variations: Among the three methods of inoculation, soil infestation and soil drenching were superior in showing root rot symptoms on inoculated plants.

Pathogenic Variability Studies: Significant difference in virulence was observed in isolates. Isolates of latter stage produced more sporulation (++++) in the medium. Similarly Desai et al. 2003 also suggested that highly virulent isolates produced abundant sporulation.

All wound inoculated plants developed dark brown lesions about the point of inoculation. Sclerotia, Pycnidiospores and mycelium of *M. phaseolina* were found in all plants 5 inoculated (Fig.1) but very less sclerotia were found in 15 days old plants and No sclerotia were found in 10 days old plants. No discoloration developed on controls

All inoculated plants starts showing symptoms in 14 days as lesions enlarged. The youngest plants (15 days old) wilted and show 3.68% disease incidence within 7-8 days and the oldest plant (60 days old) shows 90.5% disease incidence after inoculation. The average lesions size at the time of rotting increased with increase in plant age and was related to collar region size. Table 1 revealed the influence of plant age on its susceptibility to rot. It was observed that 60 days old plants (Mature Stage) bearing mature pod was more susceptible to disease than younger plants

(15 days). Maximum disease incidence in plants occurs at maturity then seedling and flowering stages. Similar studies were done by Cheng and Tu, 1972. He found that the susceptibility of plants to *Macrophomina* increased with age.

TABLE 1. Effect of crop age on root rot disease development in moth bean (*Vigna aconitifolia*) (Jacq.) Marechal.

Crop age(days)	Percent disease incidence Mean (%)	Percent disease incidence.
15	3.68	11.06
30	10.0	18.43
45	60.0	50.77
60	90.5	100

Figure in paranthesis are angular transformed values.

S.Em. +	10.8369
CD% 5.	5.15 7.50
CV%	6.67
GM	41.04

Discussion

In this study, age of plants played a significant role in disease infection. It was observed that the younger plants at 10 and 15 days were less susceptible to infection for all the inoculations methods than plants at 60 days. This agrees with Agrios (2005), who reported that plant age is important in disease infection. He also reported that plants in their reactions (susceptibility or resistance) to disease depends largely on age and for instance infections caused by *Phytophthora*; damping off and root rots, downy mildews, bacterial blight and viral infections, The host plants are susceptible during the growth period and become resistant during the adult period but that's exception with moth bean. So depending on the particular plant-pathogen combination, the greater the no. of pathogen propagules within or near fields of host plants, the more inoculum reaches the host.

The histopathology and isolation of the fungus from collar region portions suggested that the fungus mycelium grows rapidly and is associated with discoloration. The symptoms revealed dark browning and shredding of stem, numerous black microsclerotia were observed in the root and collar regions of infected plants. Similar symptoms were recorded by Vallijos (1988). Percentage disease observed 90.5% at 60 days crop age and 3.68% at younger stages. Maximum disease incidence in plants occurs at maturity than at younger stages. This may be the effect of physiological state of plants. (Sheldrake, 1978).

Root rot development in moth bean plants is probably due to tissue disintegration, caused by enzymes and toxin activity. In plants (30-60 days) old, sclerotia may contribute to symptom development, assuming that they occur in naturally infected plants. Also, viable sclerotia of the fungus in moth bean plant debris may be a source of inoculum for stored seed or for introducing the pathogen into non-infested fields. (Banerjee et al, 1982; Pareek and Jain, 1997; Upadhyay and Pandey, 2000).

Conclusion

These studies demonstrate the role and association of plant age in the causation, development and spread of root rot diseases of moth bean. The reaction of the pathogen against the plant age helps us to draw conclusion that *Macrophomina phaseolina* was found to be more wild at maturity stage. The pathogen infects the moth plant at all ages but heavy loss observed at maturity stage in comparison to other stages.

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Photomicrographs of moth bean (*Vigna aconitifolia*)(Jacq.)Marechal.Showing Pycnidiospore

